

## **FILE DEFINITION - TEST CHANGE**

Notification Date: August 23, 2022 Effective Date: September 22, 2022

# Miscellaneous Studies Using Chromosome-Specific Probes, FISH

Test ID: MISCF

# **Explanation:**

On the effective date, the changes to specimen types listed below will take place. Please note that miscellaneous FISH testing of a hematologic nature will continue to be available under test code HEMMF.

## Specimen Required:

# Current Specimens Accepted Amniotic fluid Blood Bone marrow Chorionic villi (CVS) Lymph node Skin biopsy Tissue block or slide Tumor

New Specimens Accepted
Amniotic fluid
Blood (non-hematologic testing)
Chorionic villi
Lymph node
Skin biopsy
Tissue
Tumor

## **New Specimen Requirements:**

Provide a reason for referral with each specimen. The laboratory will not reject testing if this information is not provided, but appropriate testing and interpretation may be compromised or delayed.

Advise Express Mail or equivalent if not on courier service.

### Submit only 1 of the following specimens:

Specimen Type: Amniotic fluid

**Supplies:** Refrigerate/Ambient Shipping Box, 5 lb (T329)

Container/Tube: Amniotic fluid container

**Specimen Volume:** 20-25 mL

**Collection Instructions:** 1. Optimal timing for specimen collection is during 14 to 18 weeks of

gestation, but specimens collected at other weeks of gestation are also

accepted. Provide gestational age at the time of amniocentesis.

2. Discard the first 2 mL of amniotic fluid.

3. Place the tubes in a shipping box.

4. Fill remaining space with packing material.

Specimen Type: Blood (only accepted for Congenital/Hereditary [nonhematologic]

testing)

Container/Tube: Preferred: Yellow top (ACD)

Acceptable: Green top (heparin) or lavender top (EDTA)

**Specimen Volume:** 6 mL

**Collection Instructions:** 1. Invert several times to mix blood.

2. Send whole blood specimen in original tube. **Do not aliquot.** 

3. Other anticoagulants are not recommended and are harmful to the

viability of the cells.

Specimen Type: Chorionic villi (CVS)

Supplies: CVS Media (RPMI) and Small Dish (T095)

Container/Tube: 15-mL tube containing 15 mL of transport media

**Specimen Volume**: 20 to 25 mg

**Collection Instructions:** 1. Collect specimen by the transabdominal or transcervical method.

2. Transfer chorionic villi to a Petri dish containing transport medium (e.g.,

CVS media (RPMI)).

3. Using a stereomicroscope and sterile forceps, assess the quality and

quantity of the villi and remove any blood

clots and maternal decidua.

Specimen Type: Lymph node

Supplies: Hank's Solution (T132)

**Container/Tube:** Sterile container with sterile Hank's balanced salt solution, Ringer's solution,

or normal saline

Specimen Volume: 1 cm(3)

Specimen Type: Skin biopsy

**Supplies:** Hank's Solution (T132)

Container/Tube: Sterile container with sterile Hank's balanced salt solution, Ringer's solution, or normal

saline

**Specimen Volume:** 1-cm(3) biopsy specimen of muscle/fascia from the thigh

**Collection Instructions:** 1. Wash biopsy site with an antiseptic soap.

2. Thoroughly rinse area with sterile water.

3. Do not use alcohol or iodine preparations.

4. A local anesthetic may be used.

5. Biopsy specimens are best taken by punch biopsy to include full thickness of

dermis.

Specimen Type: Tissue

Preferred: Tissue block

Collection Instructions: Submit a formalin-fixed, paraffin-embedded (FFPE) tumor tissue block. Blocks

prepared with alternative fixation methods may be acceptable; provide fixation method

used.

Additional Information: 1. Paraffin embedded specimens can be from any anatomic location (skin, soft tissue,

lymph node, etc).

2. Bone specimens that have been decalcified will be attempted for testing, but the

success rate is approximately

50%.

Acceptable: Slides

Collection Instructions: For each probe set ordered, 4 consecutive, unstained, 5 micron-thick sections placed

on positively charged slides. Include 1 hematoxylin and eosin-stained slide for the

entire test order.

Specimen Type: Tumor

**Supplies:** Hank's Solution (T132)

Container/Tube: Sterile container with sterile Hank's balanced salt solution, Ringer's solution, or normal

saline

**Specimen Volume:** 0.5 to 3 cm(3) or larger

## **Questions**

Contact Joshua Couchene, Laboratory Technologist Resource Coordinator at 800-533-1710.